

Development of a spectrophotometric method for carmoisine determination in quality control of equipment cleaning

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Introduction

Pharmaceutical products can be contaminated by other active pharmaceutical ingredients, by cleaning agents, or by other materials. Cleaning processes in pharmaceutical industry have been regulated by the implementation of the Good Manufacturing Practice rules. Cleaning procedures include sampling and testing for allowable trace amounts of active pharmaceutical ingredients or excipients that were a part of the previous preparation^{1, 2)}.

There are two methods of sampling that are considered to be acceptable: direct surface sampling (swab method) and indirect sampling (use of rinse solutions). The acceptance limits of contaminants and residues are determined in the International Conference on Harmonisation (ICH) guidelines on their specific toxicity in tested impurities^{3, 4)}.

The suitability of the material to be used for sampling and the suitability of the sampling medium should be determined. The ability to recover samples accurately may be affected by the choice of the sampling material. A major complication when using rinse solutions is probably very dilute solutions since the quantitative content of the active substance is not always possible to determine by the available analytical methods^{1, 3)}.

Carmoisine (E 122) (Fig. 1) is a red-coloured synthetic food azo dye, a derivative of diazosulfonaphthalenes, a crystalline substance that is soluble in water. It is used to provide a certain colour for many medicines as for individual matters and mixed with other dyes⁵⁾.

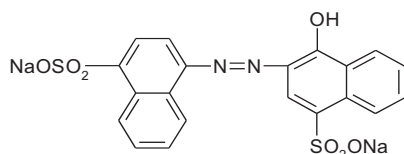


Fig. 1. Carmoisine

Methods for determining the dyes in food and medicines by HPLC are known⁶⁾. But these methods are laborious, they require a lot of time and use expensive equipment that carries great expenses to pharmaceutical manufacturers.

Experimental methods

The goal of our work is to develop an express spectrophotometric procedure for determining the residual quantities of carmoisine dye in rinse solutions to control the completeness of the process equipment cleaning.

During carrying out the experimental researches, the following materials and methods were used – the synthetic food dye carmoisine (series 657334(1-20)-657383(1-20)) and the medicinal substance myramistin (series 010514). Analytical researches were performed on a spectrophotometer Evolution 60S. For operation we used an AXIS ANG200 analytical balances, Class A measuring glassware and excipients which meet the requirements of the State Pharmacopoeia of Ukraine (SPHU)⁷⁾ which are harmonized with the European ones⁸⁾.

To develop the control methods of equipment cleaning, we propose to use an extractive photometry method using the reaction of formation of the ion associate of carmoisine with myramistin. Due to the earlier results⁹⁾, for the process of concentration of the substance contained in a large volume of liquid it is possible to use an extraction of forming ion associate by a minimum volume of an extractant. We found previously that the forming ion associate is extracted well with chloroform. We decided to use a mixture of butanol and chloroform for extraction. The sample solution was used to choose the optimal ration of chloroform and butanol.

Preparation of sample solution: 0.1 ml of $14 \cdot 10^{-3}$ M carmoisine solution was placed into a 1000.0 ml volumetric flask, 2.5 ml of $1 \cdot 10^{-3}$ M myramistin solution was added and adjusted to the mark with water R. The resulting sample solution was transferred to a separating funnel and extracted with a portion of organic solvent of 50.0 ml. The obtained extract was measured on a spectrophotometer at 523 nm wavelength against a blank solution.

The results are presented in Table 1.

One of the major requirements that determine the possibility of using spectral methods to quantify the matter is subordination of light absorption solution of Beer-Lambert law.

To test this subordination, 0.0500 g of a standard dye sample (accurately weighed) quantitatively with 60 ml of water R was transferred to a volumetric flask of 100.0 ml, adjusted to the mark with the same solvent and thoroughly mixed. The aliquots of 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0 ml, respectively, of the resulting solution were taken and placed into a 100.0 ml volumetric flask and adjusted to the mark with the same solvent. The measurement of absorbance was carried out at 517 nm wavelength in a ditch with a layer thickness of 10 mm against a blank solution.

During the control of cleaning equipment, definition of dye residues was carried in a solution with a very low concentration. We have also tested that the light absorption of carmoisine and myramistin mixture solutions subordinates the Beer-Lambert law in a concentration of $1-10 \cdot 10^{-6}$ M.

Results and discussion

Analysis of data in Table 1 shows that the optimal ratio of chloroform and butanol in the organic phase depends on the partial solubility of butanol and chloroform in water. Accordingly, it is proposed to use chloroform and butanol ratio of 1 : 1 for the extraction of the ion associate of carmoisine with miramistin from rinse solutions.

Table 1. The optimum ratio of extragents

Chloroform: butanol ratio	Absorbance
1 : 0	0.041
0 : 1	–
1 : 1	0.058
1 : 2	0.053
2 : 1	0.042
1 : 3	0.056
3 : 1	0.050

The results suggest that the solution of carmoisine subordinated the Beer-Lambert law in concentrations of 0.5–5.0 mg/100 ml. The specific absorbance is 446 ± 3 .

The results suggest that the solution of the associate subordinated the Beer-Lambert law in concentrations $1 \cdot 10^{-6} - 10 \cdot 10^{-6}$ M. The molar absorptivity is 11214 ± 460 .

Method of determination the carmoisine in quality control of equipment cleaning

Place a 1000.0 ml of rinse solution into a separating funnel, add 2.5 ml of $1 \cdot 10^{-3}$ M myramistin solution and mix. Add 50.0 ml of a mixture solution of chloroform and butanol (in the ratio 1 : 1) to the obtained solution, shake it intensively for 2 minutes and leave for 5 minutes to separate the layers. Measure the absorbance on a spectrophotometer at 523 nm wavelength against blank solution, which is used as a mixture of chloroform with butanol.

Preparing the solution of dye standard sample

0.0500 g of standard dye sample (accurately weighed) is quantitatively transferred to a volumetric flask of 100.0 ml, adjusted to the mark with the water R and thoroughly mixed. The aliquot of 1.0 ml of the resulting solution is taken and placed in a 100.0 ml volumetric flask and adjusted to the mark with the same solvent.

Calculate the residual quantities of carmoisine in rinse solution (in mg) using the formula:

$$X = \frac{A \cdot C_{st} \cdot V_{vf}}{A_{st}}$$

where absorbance of test solution, A_{st} absorbance of

standard solution, C_{st} concentration of standard solution, mg/ml.

In order to obtain the metrological characteristics of the method developed for the determination of residual quantities of carmoisine dye in rinse solutions, we used the method of modelling samples.

Preparation of sample solution: 0.1 ml of $14 \cdot 10^{-3}$ M carmoisine solution was placed into a 1000.0 ml volumetric flask, 2.5 ml of $1 \cdot 10^{-3}$ M myramistin solution was added and adjusted to the mark with water R. The resulting sample solution was transferred to a separating funnel and extraction was performed by the developed method described above.

The results are given in Table 2.

Table 2. The metrological characteristics of the developed method ($P, \% = 95, n = 6$)

ν	\bar{x}	S^2	S	$S_{\bar{x}}$	$\Delta_{\bar{x}}$	Δ_x	$\epsilon_{\bar{x}}$
5	0.00191	0.02220	0.14899	0.06078	0.38299	0.06385	8.19

Conclusions

1. A method of determination of residual quantities of carmoisine in rinse solutions from the surface of the pharmaceutical equipment was developed.

2. A simple and expressive procedure, which is characterized by satisfactory metrological characteristics, was proposed.

3. The method can be recommended for the determination of residual quantities in the control of fullness cleaning of process equipment for pharmaceutical companies.

Conflict of interest: none.

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